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Evaluation of Backcrossed Pyramiding Lines of the Yield-related Gene and the Bacterial Leaf Blight Resistant Genes

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Abstract. The yield-related gene, *WFP* increases grain number by increasing the primary branch number per panicle in rice. In the present study, *WFP* was introgressed from ST-12 to IRBB60, a pyramiding line having the genes *Xa4*, *xa5*, *xa13* and *Xa21* for BLB resistance in the IR24 genetic background. The pyramided lines PL-1, PL-4 and PL-5 that were included in the initial BC₂F₂ selections based on their improved PBN were selected for further agronomic evaluation and generation advance up to BC₂F₃. All three lines recorded significantly higher PBN and GN due *WFP* introgression, although the loss of at least two BLB genes during the breeding process resulted in the variable response of the lines to the different BLB races. Among the lines, only PL-5 showed a significantly higher estimate of actual yield measured in terms of panicle weight per square meter compared to IRBB60. PL-5 also exhibited resistance to five out of the six BLB races used for resistance screening. Despite the negative effects in grain size, the improved yield estimates, as well as the validated BLB resistance of PL-5 makes it a suitable candidate for cultivar adoption under the tropical rice ecosystem of Southeast Asia.

Key words: Rice, Wealthy Farmer's Panicle (WFP), bacterial leaf blight (BLB), pyramiding, high-yielding

Introduction

By 2050, the current world population is expected to reach 9.7 billion (https://www.un.org/development/desa/ en/). An increase of this magnitude poses a very serious threat to the current state of global food security (http:// www.fao.org/home/en/). To feed 9 billion people, global cereal production alone needs to increase by 38% in the

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next 34 years1).

Rice, wheat and maize constitute approximately 50% of the calorie source of the human diet²). Among these three, rice serves as a staple for more than half of the world's population. Although rice is cultivated in more than 100 countries across the globe, more than 90% of rice is still produced and consumed in Asia (GRiSP, http:// www.grisp.net/main/summary). In recent years, a drastic increase in demands for rice has also been reported in Africa³ (GRiSP, http://www.grisp.net/main/summary). Given the importance of rice as a staple food, increasing

rice production would play a critical role in securing world food supply.

Marker-assisted selection (MAS) is powerful tool for an efficient breeding. Since MAS makes it possible to distinguish plants into which target genes/QTLs have been introgressed during nursery, field area and efforts for individual selection of breeding reduce. Also, target genes/ QTLs can be introgressed more reliably than visually selection. MAS has been successfully used in breeding programs to improve agronomic traits, as well as biotic and abiotic stress resistance in rice. In the past decades, MAS have significantly shortened the time requirement while increasing the precision and efficacy by which target genes/QTLs are transferred across different genetic backgrounds^{4, 5)}. To date, several rice cultivars have been improved through MAS and these include the improved varieties developed under the Wonder Rice Initiative for Food Security and Health (WISH) project (http:// motoashikari-lab.com/wp/wp-content/uploads/2018/11/ WISH catalogue lite4.pdf).

To effectively utilize MAS in breeding programs, genes/QTLs underlying important traits need to be mapped and identified. In rice, many genes/QTLs that are related to the sink ability have been identified including *GN1a/OsCKX2*⁶, *WFP/IPA1*^{7,8}, *NAL1/SPIKE/GPS*^{9,10,11}, *APO1/SCM2*^{12,13,14}, *qSW2*¹⁵, *GW2*¹⁶, *GW6a*¹⁷) and *GS3*^{18,19}. Conversely, several genes/QTLs that are related to the source or translocation ability of rice has also been reported^{20, 21, 22} that includes *AMY2A*²³, *BAM2*²³ and *GWD1*²⁴.

Wealthy Farmer's Panicle (WFP)/Ideal Plant Architecture 1 (IPA1) is a yield-related gene that encodes the plant-specific transcription factor OsSPL14^{7,8}). OsSPL14 contains a microRNA-targeted sequence in an exon that is targeted and cleaved by OsmiR156. OsSPL14 functions in the regulation of primary branch number and is therefore associated with grain number. The WFP allele from ST-12 has been reported to increase grain number per panicle in rice⁷). Advanced breeding lines of the *japonica* rice cultivar Nipponbare and several other *indica* cultivars having the WFP allele from ST-12 showed at least 28% increase in grain yield compared to the original varieties^{7, 25}).

Bacterial leaf blight (BLB) caused by *Xanthomonas* oryzae pv. oryzae (Xoo) is one of the most serious diseases of rice throughout the world. In some regions in Asia, BLB can cause yield losses of up to 50%²⁶). The most effective approach to combat BLB is the use of resistant cultivars²⁷). To date, many resistant genes against BLB have been identified. Huang et al. focused on four BLB resistance genes/QTLs namely *Xa4*, *xa5*, *xa13* and *Xa21*, and pyramided all four resistance genes in the IR24 genetic background to generate IRBB60²⁸). IRBB60 has resistance

to six races of *Xoo* from the Philippines²⁸⁾. Currently, the BLB genes xa5, xa13 and Xa21 have been identified and characterized^{29, 30, 31)}.

In this study, we aimed to pyramid *WFP* from ST-12 to IRBB60, a rice cultivar having four genes for BLB resistance. The resulting pyramided lines were evaluated for yield and yield components, as well as for their resistance to BLB races from Vietnam, Myanmar and India. Results of the preliminary assessment of the agronomic performance of advanced backcrosses lines identified PL-5, a pyramiding line with higher yield compared and comparative BLB resistance relative to the recipient parent IRBB60.

Materials and Methods

Marker-assisted introgression of WFP in the pyramiding line IRBB60

IRBB60, IR24, ST-12 and three pyramiding lines (PL-1, PL-4 and PL-5) at the BC₂F₄ generation were used in the study. IRBB60 is a pyramiding line having the four bacterial leaf blight (BLB) resistant genes, Xa4, xa5, xa13 and Xa21 in the genetic background of IR24²⁸), whereas ST-12 is the donor of the yield-related gene, WFP7). Phenotype and marker-assisted backcrossing of the initial hybrids generated from crosses between IRBB60 and ST-12 was carried out up to the BC₂F₄ generation (Fig. 1). An SSR marker (forward primer: GCGGTAACAAACCAACCAACC, reverse primer: AAAGCAGGACACAGTCACAGG) was used to monitor the introgression of WFP in the IRBB60 background. In 2016, six BC₂F₃ lines (PL-1, PL-2, PL-4, PL-5, PL-6 and PL-7) were selected for preliminary evaluation of yield and yield components in the field. Based on this preliminary investigation, three pyramiding lines namely PL-1, PL-4 and PL-5 were selected and advanced for further performance evaluation in the field.

All test materials were raised in the greenhouse for four weeks before transplanting them in a rice paddy at the Togo Field Center for Research and Education of Nagoya University in Aichi, Japan (35°06' north, 137°04' east) in 2017. The seedlings were transplanted separately in 3 rows (1.2 m) at a density of 22.2 plants/m² and at a plant spacing of 30cm x 15cm. All materials were grown in three replications in randomized blocks and under low fertilizer conditions i.e. 300 kg K-P-N (16-12-14)/ha only before transplanting.

Evaluation of the pyramiding lines for agronomic traits

Before sampling panicles from all materials, data on the days to heading (DH), plant height (PH) and panicle length (PL) of five plants per block were recorded from





plant stands in the paddy. At 40-45 days after heading, all panicles from five plants from each block were sampled. After sampling, panicle weight per plant (PW), panicle number per plant (PN), primary branch number per main panicle (PBN) and grain number per main panicle (GN) were measured. To estimate the actual yield of each pyramiding line in the field, panicle weight per plant was measured and converted to panicle weight per square meter (PWm²). Only plants within rows were sampled to avoid border effects. All panicles were dried for a week in an incubator set at 42°C. To determine fertility ratio (FR), the three largest panicles including the main panicle were chosen from each plant. FR was calculated by dividing the number of filled spikelets by the total number of spikelets, multiplied by 100. The grain weight and number of filled spikelets of the three largest panicles were measured and converted to 1000-grains weight per main three panicles (TGW). Grain shape based on grain length (GL), width (GW) and thickness (GT) was determined from ten randomly selected spikelets in each plant. For the statistical analyses, plants showing maximum and minimum PW in each plot were excluded.

Evaluation of the pyramided lines for bacterial leaf blight resistance

Six races of Xanthomonas oryzae pv. oryzae (Xoo) causing bacterial leaf blight (BLB) from Vietnam (HAU01043, HAU0209, HAU02021-2 and HAU02024-6)³²⁾, Myanmar (HKM13)³³⁾ and India (IND2-3) were used to evaluate the resistance of the experimental materials to the pathogen. At booting stage, the plants were inoculated with *Xoo* using the leaf clipping method³⁴. A preliminary test was conducted using IR24 to confirm the pathogenicity and virulence of the six BLB races. Immediately after that, all the experimental materials including IR24 as the susceptible control and IRBB60 as the resistant control, were screened against the pathogen. After two weeks from inoculation, lesion lengths were measured, and the resistance/susceptibility of each material was evaluated. Classification of the response of materials as resistant, moderate, susceptible or partially resistant was carried out based on a comparison to the response of the susceptible reference IR24 and the resistant reference IRBB60 using Tukey's HSD (Table 1). Screening for BLB resistance including maintenance and culture of the pathogen was

Table 1. Classification of the response of materials to the BLB races based on acomparison to the response of the susceptible reference IR24 (S) andthe resistant reference IRBB60 (R).

Statistical difference to IR24 (S)	Statistical difference to IRBB60 (R)	Response	
Significant*	Not significant	Resistant	
Significant	Significant	Moderate	
Not significant	Significant	Susceptible	
Not significant	Not significant	Partially resistant**	

*; significant according to Tukey's HSD (p < 0.05).

**; only in the case that evaluation value is between the values of the references.

carried out in Laboratory of Plant Pathology, Faculty of Agriculture, Kyushu University.

Genotyping-by-sequencing analysis of the pyramiding lines

Genotyping-by-sequencing (GBS) was conducted for the three pyramiding lines at the BC₂F₄ generation using the Illumina Miseq sequencing system and Illumina Miseq Reagent Kit v3 (150 cycles)^{35, 36)}. Three plants from each of the pyramiding line, as well as IRBB60 and ST-12 were genotyped by GBS for whole genome. After sequencing, the reads were trimmed, and genotypes were called and filtered using TASSEL-GBS³⁷⁾. SNPs with minor allele frequency (MAF) of >0.02, as well as lines with >0.2 lost genotype data and with >0.125 degree of heterozygous sites were removed from genotyping. Finally, imputation and error correction were carried out with FSFHap in TASSEL 4.0³⁸).

Statistical analysis

T-test and Tukey's HSD were performed using the free statistical software R (https://www.r-project.org/). A significant difference in Tukey's HSD was set at p<0.05. R was also used for the randomization of blocks in the field design.

Result

Phenotypic evaluation of the three pyramiding lines for yield components

The pyramiding lines PL-1, PL-2, PL-4, PL-5, PL-6 and PL-7 in the genetic background of IRBB60 were selected visually from a BC_2F_2 population for their high primary branching number. The introgression of *WFP* from ST-12 in these six lines at the BC_2F_3 generation was confirmed by MAS. Evaluation for agronomic and yield-related traits showed that except for PL-6, all the pyramiding lines had

significantly more PBN than IRBB60 (Table 2). PL-6 and PL-7 recorded significant lower PN (panicle number per plant) than IRBB60. The reduction of PN is the negative effect for actual yield, hence both PL-6 and PL-7 were removed from the candidate lines. Similarly, PL-2 was removed from the candidate lines because of comprehensive gross morphology. Based on the overall agronomic performance and gross morphology of the pyramiding lines, we selected PL-1, PL-4 and PL-5 for further generation advance and agronomic evaluation (Fig. 2).

At the BC₂F₄ generation, PL-1, PL-4 and PL-5 were evaluated for yield components including DH, PH, PL, PN, GN, PBN, FR, TGW, GL, GW and GT (Table 3). Compared to IRBB60, PL-1 recorded a significant 4-day delay in a heading, whereas PL-5 was significantly early by 4 days. Heading date in PL-4 was equivalent to that of IRBB60. PN decreased by 0.33 to 1.33 in all pyramiding lines compared to IRBB60. GN (grain number per main panicle) and PBN (primary branch number per main panicle) of the three pyramiding lines were significantly higher than those of IRBB60, suggesting the positive effects of WFP introgression from ST-12. The PBN of ST-12 was twice higher than that of IRBB60 and IR24. FRs (fertility ratio per three main panicles) of IRBB60, IR24 and PL-5 showed high values, while those of ST-12, PL-1 and PL-4 were low, below 77%. Among the three pyramiding lines, PL-5 showed the highest FR (92.56 \pm 0.40%). The FRs of PL-1 and PL-4 were 76.84 \pm 2.37% and $72.64 \pm 0.61\%$ respectively, which were equivalent to that of ST-12.

In PWm², PL-5 recorded a significantly higher yield (845.50 \pm 57.20 g/m²) than IRBB60 (782.42 \pm 47.90 g/m²), with the rate of increase in PL-5 reaching up to 8.1% of IRBB60. ST-12, the donor of *WFP*, had significantly lower PWm² (712.4 \pm 43.5 g/m²) than IRBB60 (Fig. 3). The panicle weight of PL-1 and PL-4 were statistically equivalent to that of IRBB60.

Table 2. Yield-components of the pyramiding lines at BC₂F₃ in the IRBB60 background.

	PH (cm)	PL (cm)	PN	GN	PBN
IRBB60	97.00 ± 4.36 ac*	$26.40\pm0.51~ab$	11.33 ± 0.58 a	203.33 ± 17.79 a	12.33 ± 0.58 a
ST-12	$115.00\pm7.55~bd$	$25.43\pm1.33~b$	$7.00\pm1.00\ bc$	$280.67 \pm 43.35 \ ab$	$21.00\pm4.00\ bc$
PL-1	119.33 ± 3.21 b	29.26 ± 0.61 a	8.67 ± 0.85 ac	314.67 ± 66.83 ab	23.33 ± 1.53 b
PL-2	$98.50\pm 6.36~ac$	$25.48\pm1.52~ab$	11.00 ± 1.41 ad	$281.00 \pm 31.11 \ ab$	$23.00\pm1.41\ b$
PL-4	$114.33\pm2.52~bd$	$26.40\pm1.03~ab$	11.33 ± 1.53 a	312.67 ± 33.56 ab	$24.67\pm0.58~b$
PL-5	$102.33\pm5.13\ \text{acd}$	$23.43\pm0.99\ ab$	$8.33 \pm 1.53 \text{ ab}$	$274.33 \pm 15.53 \ ab$	$20.33\pm1.53~bc$
PL-6	$89.33\pm6.43~a$	$24.90\pm1.73~ab$	$5.33\pm0.58~b$	$204.00 \pm 33.05 \ a$	17.00 ± 1.00 ac
PL-7	103.67 ± 1.15 cd	$25.30\pm1.63~b$	7.67 ± 1.53 bcd	354.33 ± 62.01 b	$21.00\pm1.00\ bc$

Numbers represent average \pm standard deviation.

PH=plant height; PL=panicle length; PN=panicle number per plant; GN=grain number per main panicle; PBN=primary branch number per main panicle.

*; different letters indicate significant differences at p < 0.05 by Tukey's HSD.



Fig. 2. Gross morphology of the plants and panicles of the experimental materials. A-F: Gross plant morphology at the ripening stage, G-L: Gross panicle morphology at the ripening stage. A and G: IRBB60, B and H: IR24, C and I: ST-12, D and J: PL-1, E and K: PL-4 and F and L: PL-5. Bar = 20cm (A–F), 5cm (G–L).

Table 3. Yield-components of the pyramiding lines at BC_2F_4 in the IRBB60 background.

DH (days)	PH (cm)	PL (cm)	PN	GN	PBN
105.00 ± 0.50	94.67 ± 1.55	23.89 ± 0.44	10.44 ± 0.65	160.22 ± 9.47	12.00 ± 0.37
$109.44 \pm 0.33^{**}$	$83.89 \pm 1.38 **$	$22.22 \pm 0.58 **$	10.33 ± 0.80	161.33 ± 10.07	$12.78\pm0.37\texttt{*}$
$95.78 \pm 0.33 **$	$101.89 \pm 1.14 \texttt{**}$	23.78 ± 0.53	$8.89 \pm 0.50 **$	$283.11 \pm 15.61 ^{\ast\ast}$	$22.33 \pm 0.96 \text{**}$
$109.11 \pm 0.44 ^{**}$	$103.33 \pm 2.39 **$	$25.78\pm1.14\texttt{*}$	9.11 ± 0.93	$219.89 \pm 16.68 ^{\ast\ast}$	$20.11 \pm 0.65 **$
105.22 ± 0.53	96.67 ± 1.98	23.22 ± 0.50	$9.44\pm0.41\texttt{*}$	$231.78 \pm 10.14 \texttt{**}$	$21.44 \pm 1.40 **$
$101.11 \pm 0.67 \textit{**}$	95.89 ± 1.71	$22.00 \pm 0.24 \text{**}$	10.11 ± 0.80	$239.89 \pm 9.29 \texttt{**}$	$20.56 \pm 0.75^{\ast\ast}$
FR (%)	TGW (g)	GL (mm)	GW (mm)	GT (mm)	
94.90 ± 0.26	26.45 ± 0.08	9.46 ± 0.02	2.41 ± 0.01	1.96 ± 0.01	
$88.14 \pm 0.79 **$	$25.65 \pm 0.11 **$	$9.11 \pm 0.03 **$	2.52 ± 0.01 **	$1.97\pm0.01*$	
$72.75 \pm 1.14 **$	$23.73 \pm 0.11 **$	$9.87 \pm 0.02 **$	2.33 ± 0.01 **	$1.94 \pm 0.01 **$	
$76.84 \pm 2.37 **$	$27.55 \pm 0.40 \text{**}$	$9.56 \pm 0.06 **$	2.45 ± 0.02 **	$1.91 \pm 0.02 **$	
$72.64 \pm 0.61 **$	$24.40 \pm 0.09 **$	9.47 ± 0.02	2.41 ± 0.01	1.86 ± 0.01 **	
$92.56 \pm 0.40 \text{**}$	20.83 ± 0.11 **	8.60 ± 0.01 **	2.22 ± 0.01 **	$1.79 \pm 0.01 ^{**}$	
	$\begin{array}{c} DH \ (days) \\ 105.00 \pm 0.50 \\ 109.44 \pm 0.33^{**} \\ 95.78 \pm 0.33^{**} \\ 109.11 \pm 0.44^{**} \\ 105.22 \pm 0.53 \\ 101.11 \pm 0.67^{**} \\ \hline \\ FR \ (\%) \\ 94.90 \pm 0.26 \\ 88.14 \pm 0.79^{**} \\ 72.75 \pm 1.14^{**} \\ 76.84 \pm 2.37^{**} \\ 72.64 \pm 0.61^{**} \\ 92.56 \pm 0.40^{**} \\ \end{array}$	$\begin{array}{c c} DH \ (days) & PH \ (cm) \\ \hline 105.00 \pm 0.50 & 94.67 \pm 1.55 \\ 109.44 \pm 0.33^{**} & 83.89 \pm 1.38^{**} \\ 95.78 \pm 0.33^{**} & 101.89 \pm 1.14^{**} \\ 109.11 \pm 0.44^{**} & 103.33 \pm 2.39^{**} \\ 105.22 \pm 0.53 & 96.67 \pm 1.98 \\ 101.11 \pm 0.67^{**} & 95.89 \pm 1.71 \\ \hline \\ \hline \\ FR \ (\%) & TGW \ (g) \\ \hline \\ 94.90 \pm 0.26 & 26.45 \pm 0.08 \\ 88.14 \pm 0.79^{**} & 25.65 \pm 0.11^{**} \\ 72.75 \pm 1.14^{**} & 23.73 \pm 0.11^{**} \\ 76.84 \pm 2.37^{**} & 27.55 \pm 0.40^{**} \\ 72.64 \pm 0.61^{**} & 24.40 \pm 0.09^{**} \\ 92.56 \pm 0.40^{**} & 20.83 \pm 0.11^{**} \\ \hline \end{array}$	$\begin{array}{c c c c c c c c c c c c c c c c c c c $	$\begin{array}{c c c c c c c c c c c c c c c c c c c $	$\begin{array}{c c c c c c c c c c c c c c c c c c c $

Numbers represent average \pm standard error.

DH=days to heading; PH=plant height; PL=panicle length; PN=panicle number per plant; GN=grain number per main panicle; PBN=primary branch number per main panicle; FR=fertility ratio per three largest panicles in a plant; TGW=1000-grain weight per three largest panicles in a plant; GL=grain length; GW=grain width; GT=grain thickness.

*, **; indicate significant difference from IRBB60 at p<0.05 and p<0.01 by t-test, respectively.

Based on these results, the high yield of PL-5 can be attributed to the increase in GN and PBN. In spite of PL-5 having 49.7% higher GN than IRBB60, PWm² of PL-5 was only 8.1% higher. As a major cause of this phenomenon, we focused on grain size. The grain size is expected to affect TGW (1000-grains weight per main three panicles). Compared to the TGW of IRBB60 (26.45 ± 0.08 g), the TGW of PL-5 (20.83 ± 0.11 g) was significantly lower (Table 3). Similarly, PL-5 recorded smaller grains in terms of GL (8.60 ± 0.01 mm), GW (2.22 ± 0.01 mm) and GT (1.79 ± 0.01 mm) compared to those of IRBB60 ($9.46 \pm$ 0.02 mm, $2.41 \pm 0.01 \text{ mm}$, $1.96 \pm 0.01 \text{ mm}$) (Table 3). The smaller grains of PL-5 negatively affected its yield.

Evaluation of the pyramiding line for resistance to bacterial leaf blight

The response of the experimental materials when challenged with BLB races from Vietnam (HAU01043, HAU0209, HAU02021-2 and HAU02024-6), Myanmar (HKM13) and India (IND2-3) are shown in Fig. 4. IRBB60 showed strong resistance to the six BLB races, with lesions lengths ranging from 0 to 1.2 cm. In contrast,



Fig. 3. Panicle weight per square meter (PWm²) of the experimental materials. * and ** indicate significant difference from IRBB60 at p<0.05 and p<0.01, respectively, by t-test. Error bars show standard error.

IR24 and ST-12 were susceptible to the six BLB races, with lesions lengths ranging from 13.3 to 25.2 cm in IR24 and from 10.5 to 26.8 cm in ST-12. The pyramiding lines PL-1, PL-4 and PL-5 showed differential responses to the six BLB races. PL-1 exhibited resistance to the five BLB races from Vietnam (HAU0209, HAU02021-2 and HAU02024-6), Myanmar (HKM13) and India (IND2-3), and moderate to HAU01043. PL-4 exhibited resistance to HKM13 and susceptibility to HAU01043, HAU02021-2 and HAU02024-6. PL-5 exhibited resistance to the five BLB races from Vietnam (HAU01043, HAU02021-2 and HAU02024-6. PL-5 exhibited resistance to the five BLB races from Vietnam (HAU01043, HAU02021-2 and HAU02024-6) and India (IND2-3), and partially resistance to HKM13 from Myanmar.

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GBS analysis validated the introduction of the *WFP* allele from ST-12 and the segregation of the four BLB resistant genes of IRBB60 in each of the pyramiding lines at BC_2F_3 (Table 4). All pyramiding lines carry the *Xa21* allele from IRBB60 and the *xa13* allele from ST-12. PL-1 and PL-4 have the *Xa4* allele from IRBB60 but not PL-5 which has the *Xa4* from ST-12. The *xa5* allele from IRBB60 was introgressed in the homozygous form in PL-5 but occurs in a heterozygous form in PL-4. PL-1 has the ST-12 allele for *xa5*.

Discussion

The pyramiding lines (BC_2F_4) reported in the present study were developed from the cross between ST-12 (used as the donor of WFP) and IRBB60 (used as the recipient parent with BLB resistance). Introgression of WFP from ST-12 in the PLs was confirmed by MAS and GBS (Fig. 1, Table 4). As a result, we developed PL-5, a pyramiding line with higher yield compared to the recipient parent, IRBB60 (Fig. 3). The high yield of PL-5 can be attributed to the increase in both GN and PBN (Table 2, Table 3) due to the introgression of WFP from ST-12. Miura et al. reported that the introgression of WFP from ST-12 can significantly increase both GN and PBN in lines having the genetic background of the japonica rice cultivar Nipponbare7). Similarly, introgression of WFP has been reported to increase GN and PBN of several rice cultivars with indica genetic backgrounds²⁵⁾. In the present study, the positive effects of WFP from ST-12 was also observed in the genetic background of IRBB60, suggesting that WFP expression can increase GN and PBN in both japonica and indica rice backgrounds, and that WFP from ST-12 is a useful genetic resource that can be used to significantly improve the yield of rice.

PL-5 recorded 49.73% higher GN and 71.33% higher PBN compared to IRBB60. Despite the significant increase in GN and PBN, PL-5 recorded a remarkably low increase

	Chromosome	RAP-ID	QTL location*	IRBB60	ST-12	PL-1	PL-4	PL-5	Exp**
Xa4	11	-	Around 28.5Mb	А	В	А	А	В	А
xa5	5	Os05g0107700	-	А	В	В	Η	А	А
xa13	8	Os08g0535200	-	А	В	В	В	В	А
Xa21	11	Os11g0559200	-	А	В	А	А	А	А
WFP	8	Os08g0509600	-	А	В	В	В	В	В

*; physical distance showed in RAP-db site (https://rapdb.dna.affrc.go.jp/).

**; expected genotypes.



Fig. 4. The response of the experimental materials to the six races of Xoo. A: Lesion length on leaves of the experimental materials. Different letters indicate significance at p<0.05 by Tukey's HSD. Error bars show standard error. B: Lesions in leaves infected by the six BLB races. The reaction of the experimental materials to the BLB races is shown in two leaves each. The order of BLB races in each panel is HAU01043, HAU0209, HAU02021-2, HAU02024-6, HKM13 and IND2-3 from left to right. The inoculation site corresponds to the top of each panel. Bars = 5cm.</p>

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of 8.06% in yield (PWm²) compared to IRBB60. A similar phenomenon was observed in PL-1 and PL-4. Rice yield has four important components namely PN, GN, FR and TGW. In PL-5, the main negative factor was TGW, which was 21.25% lower than IRBB60. Based on the results of the current study, an improvement in TGW can further increase rice yield. In PL-1, the main negative factors were PN and FR which were 12.74% and 19.03% lower than IRBB60, respectively. In PL-4, the negative factors were PN, FR and TGW which were 9.58%, 23.46% and 7.75% lower than IRBB60, respectively.

The low TGW of PL-5 may be unrelated to the function of *WFP*. PL-5 has smaller grains, giving it lower TGW than IRBB60. The donor parent ST-12 do not have small grains, and *WFP/IPA1* from ST-12 has not been reported to be involved in the regulation of grain size. We hypothesize that the genetic background of PL-5 which resulted from random recombination between the ST-12 and IRBB60 genome contributed to its smaller grains.

Miura et al. suggested that *WFP* from ST-12 is involved in the regulation of shoot branching in the vegetative stage⁷). It was also suggested that *WFP/IPA1* from the *japonica* cultivar Aikawa 1 and from the *japonica* cultivar Shaoniejing reduces tiller number^{7, 8}). While the presence of *WFP/IPA1* allele may increase GN, it may have negative effects on PN. In the present study, an increase in GN due to *WFP/IPA1* introgression resulted in a concomitant decrease in PN. To suppress any reduction in PN due to *WFP* introgression, it is necessary to modify factor(s) related to controlling PN specifically in the vegetative stage.

It is considered that the low FR of PL-1 and PL-4 occurred as a result of the inability of the source to keep up with the improved sink ability of both lines as conferred by *WFP*. In PL-5, a remarkable reduction in grain size has been confirmed that allowed the source to cope with the increase in sink. In the context of sink-source relationship, *WFP* is a sink ability-related gene. Improvement of the sink is necessary to improve yield, but this requires a concomitant improvement in the source³⁹). Efficient breeding for high yield in rice requires elucidation of the genetic factors regulating the ability of the source to generate photosynthetic assimilates that will fill the sink. Improving the photosynthetic ability of plant tissues with source functions may be the key to breaking through the yield potential of the current high-yield varieties⁴⁰).

The present study aimed to pyramid *WFP* with four BLB resistant genes already present in the IRBB60 background. The three selected pyramiding lines however, had *WFP* and at least two BLB resistant genes each (Fig. 4, Table 4). The incomplete maintenance of all four BLB resistance genes in the pyramiding line accounts for the variability in the response of each line to each of the BLB races. While

the BLB resistant genes are not linked, *xa13* is linked to *WFP* at approximately 1.5 Mb distance downstream. In the process of developing these pyramiding lines, genotyping was carried out to monitor the introgression of only *WFP*. To ensure that all four BLB resistant genes are maintained in the pyramided line, these genes should also be monitored in the backcrossed and selfed generations by MAS.

Despite losing at least two BLB resistant genes, PL-5 remains resistant to four BLB races from Vietnam and a race from India, whereas PL-1 is resistant to three races from Vietnam and races from Myanmar and India. These results indicate the both PL-1 and PL-5 can be cultivated in areas with a reported incidence of the specific BLB infection. Overall, the significantly improved yield of PL-5 in combination with its resistance to five known BLB races from Vietnam and India makes this pyramiding line a viable candidate for high yield line with adoption in tropical rice ecosystems in Southeast Asia.

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References

- 1. Tester M, Langridge P. (2010) Breeding technologies to increase crop production in a changing world. Science 327: 818-822.
- Khush G. (2003) Productivity improvements in rice. Nutr Rev 61: S114-S116.
- Seck PA, Diagne A, Mohanty S, Wopereis MCS. (2012) Crops that feed the world 7: rice. Food Security 4: 7-24.
- Angeles-Shim RB, Ashikari M. (2017) Advances in molecular breeding techniques for rice. In: Achieving sustainable cultivation of rice Vol 1. Burleigh Dodds Science Publishing, UK: 27-49.
- Kurokawa Y, Noda T, Yamagata Y, Angeles-Shim R, Sunohara H, Uehara K, Furuta T, Nagai K, Jena K, Yasui H, Yoshimura A, Ashikari M, Doi K. (2016) Construction of a versatile SNP array for pyramiding useful genes of rice. Plant Sci 242: 131-139.
- Ashikari M, Sakakibara H, Lin S, Yamamoto T, Takashi T, Nishimura A, Angeles ER, Qian Q, Kitano H, Matsuoka M. (2005) Cytokinin oxidase regulates rice grain production. Science 309: 741-745.
- Miura K, Ikeda M, Matsubara A, Song XJ, Ito M, Asano K, Matsuoka M, Kitano H, Ashikari M. (2010) OsSPL14 promotes panicle branching and higher

grain productivity in rice. Nat Genet 42: 545-549.

- Jiao Y, Wang Y, Xue D, Wang J, Yan M, Liu G, Dong G, Zeng D, Lu Z, Zhu X, Qian Q, Li J. (2010) Regulation of *OsSPL14* by OsmiR156 defines ideal plant architecture in rice. Nat Genet 42: 541-544.
- Qi J, Qian Q, Bu Q, Li S, Chen Q, Sun J, Liang W, Zhou Y, Chu C, Li X, Ren F, Palme K, Zhao B, Chen J, Chen M, Li C. (2008) Mutation of the rice narrow leaf1 gene, which encodes a novel protein, affects vein patterning and polar auxin transport. Plant Physiol 147: 1947-1959.
- Fujita D, Trijatmiko KR, Tagle AG, Sapasap MV, Koide Y, Sasaki K, Tsakirpaloglou N, Gannaban R, Nishimura T, Yanagihara S, Fukuta Y, Koshiba T, Slamet-Loedin I, Ishimaru T, Kobayashi N. (2013) *NAL1* allele from a rice landrace greatly increases yield in modern indica cultivars. Proc Natl Acad Sci 110: 20431-20436.
- 11. Takai T, Adachi S, Taguchi-Shiobara F, Sanoh-Arai Y, Iwasawa N, Yoshinaga S, Hirose S, Taniguchi Y, Yamanouchi U, Wu J, Matsumoto T, Sugimoto K, Kondo K, Ikka T, Ando T, Kono I, Ito S, Shomura A, Ookawa T, Hirasawa T, Yano M, Kondo M, Yamamoto T. (2013) A natural variant of *NAL1*, selected in high-yield rice breeding programs, pleiotropically increases photosynthesis rate. Sci Rep 3. doi: 10.1038/ srep02149.
- 12. Ikeda K, Nagasawa N, Nagoto Y. (2005) *ABERRANT PANICLE ORGANIZATION 1* temporally regulates meristem identity in rice. Dev Bio 282: 349-360.
- Ikeda K, Ito M, Nagasawa N, Kyozuka J, Nagato Y. (2007) Rice *ABERRANT PANICLE ORGANIZATION I*, encoding an F-box protein, regulates meristem fate. Plant J 51: 1030-1040.
- Ookawa T, Hobo T, Yano M, Murata K, Ando T, Miura H, Asano K, Ochiai Y, Ikeda M, Nishitani R, Ebitani T, Ozaki H, Angeles E, Hirasawa T, Matsuoka M. (2010) New approach for rice improvement using a pleiotropic QTL gene for lodging resistance and yield. Nat Commun 1: 132.
- 15. Zhan X, Sun B, Lin Z, Gao Z, Yu P, Liu Q, Shen X, Zhang Y, Chen D, Cheng S, Cao L. (2015) Genetic mapping of a QTL controlling source--sink size and heading date in rice. Gene 571: 263-270.
- Song XJ, Huang W, Shi M, Zhu MZ, Lin HX. (2007) A QTL for rice grain width and weight encodes a previously unknown RING-type E3 ubiquitin ligase. Nat Genet 39: 623-630.
- 17. Song XJ, Kuroha T, Ayano M, Furuta T, Nagai K, Komeda N, Segami S, Miura K, Ogawa D, Kamura T, Suzuki T, Higashiyama T, Yamasaki M, Mori H, Inukai Y, Wu J, Kitano H, Sakakibara H, Jacobsen SE, Ashikari M. (2015) Rare allele of a previously unidentified histone H4 acetyltransferase enhances

grain weight, yield and plant biomass in rice. Proc Natl Acad Sci 112: 76-81.

- 18. Fan C, Xing Y, Mao H, Lu T, Han B, Xu C, Li X, Zhang Q. (2006) GS3, a major QTL for grain length and weight and minor QTL for grain width and thickness in rice, encodes a putative transmembrane protein. Theor Appl Genet 112: 1164-1171.
- Takano-Kai N, Jiang H, Kubo T, Sweeney M, Matsumoto T, Kanamori H, Padhukasahasram B, Bustamante C, Yoshimura A, Doi K, McCouch S. (2009) Evolutionary History of GS3, a Gene Conferring Grain Length in Rice. Genetics 182: 1323-1334.
- 20. Wang DR, Han R, Wolfrum EJ, McCouch SR. (2017) The buffering capacity of stems: genetic architecture of nonstructural carbohydrates in cultivated Asian rice, *Oryza sativa*. New Phytol 215: 658-671.
- 21. Zhang J, Li G, Huang Q, Liu Z, Ding C, Tang S, Chen L, Wang S, Ding Y, Zhang W. (2017) Effects of culm carbohydrate partitioning on basal stem strength in a high-yielding rice population. Crop J 5: 478-487.
- 22. Phung DH, Sugiura D, Sunohara H, Makihara D, Kondo M, Nishiuchi S, Doi K. (2019) QTL analysis for carbon assimilate translocation-related traits during maturity in rice (*Oryza sativa* L.). Breed Sci 69: 289-296.
- 23. Hirose T, Aoki N, Harada Y, Okamura M, Hashida Y, Ohsugi R, Miyao A, Hirochika H, Terao T. (2013) Disruption of a rice gene for α-glucan water dikinase, *OsGWD1*, leads to hyperaccumulation of starch in leaves but exhibits limited effects on growth. Front Plant Sci 4. doi: 10.3389/fpls.2013.00147.
- 24. Wada H, Masumoto-Kubo C, Tsutsumi K, Nonami H, Tanaka F, Okada H, Erra-Balsells R, Hiraoka K, Nakashima T, Hakata M, Morita S. (2017) Turgor-responsive starch phosphorylation in *Oryza sativa* stems: A primary event of starch degradation associated with grain-filling ability. PLoS One 12: e0181272. doi: 10.1371/journal.pone.0181272.
- 25. Kim SR, Ramos JM, Hizon RJM, Ashikari M, Virk P, Torres E, Nissila E, Jena K. (2018) Introgression of a functional epigenetic *OsSPL14 WFP* allele into elite *indica* rice genomes greatly improved panicle traits and grain yield. Sci Rep 8. doi: 10.1038/s41598-018-21355-4.
- 26. Shafinah K, Sahari N, Sulaiman R, Yusoff MSM, Ikram MM. (2013) A framework of an expert system for crop pest and disease management. J Theor Appl Info Tech 58: 182-190.
- 27. Khush GS, Mackill DJ, Sidhu GS. (1989) Breeding rice for resistance to bacterial blight. In: Bacterial blight of rice. International Rice Research Institute, Manila, Philippines: 207-217.
- 28. Huang N, Angeles ER, Domingo J, Magpantay G, Singh S, Zhang G, Kumaravadivel N, Bennett J,

JISNA

Khush G. (1997) Pyramiding of bacterial blight resistance genes in rice: marker-assisted selection using RFLP and PCR. Theor Appl Genet 95: 313-320.

- 29. Jiang GH, Xia ZH, Zhou YL, Wan J, Li DY, Chen RS, Zhai WX, Zhu LH. (2006) Testifying the rice bacterial blight resistance gene *xa5* by genetic complementation and further analyzing *xa5* (*Xa5*) in comparison with its homolog *TFILA*γ*1*. Mol Gen Genom 275: 354-366.
- Antony G, Zhou J, Huang S, Li T, Liu B, White F, Yang B. (2010) Rice *xa13* recessive resistance to bacterial blight is defeated by induction of the disease susceptibility gene Os-*11N3*. Plant Cell 22: 3864-3876.
- 31. Song W, Wang G, Chen L, Kim H, Pi L, Holsten T, Gardner J, Wang B, Zhai W, Zhu L, Fauquet C, Ronald P. (1995) A receptor kinase-like protein encoded by the rice disease resistance gene, *Xa21*. Science 270: 1804-1806.
- Furuya N, Taura S, Goto T, Thuy BT, Ton PH, Tsuchiya K, Yoshimura A. (2012) Diversity in virulence of *Xan-thomonas oryzae* pv. *oryzae* from Northern Vietnam. Jpn Agricul Res Quarterly 46: 329-338.
- 33. Aye SS, Matsumoto M, Kaku H, Goto T, Furuya N, Yoshimura A. (2007) Evaluation of Resistance in Rice Plants to Myanmar Isolates of *Xanthomonas oryzae* pv. *oryzae*. J Fac Agr Kyushu Univ 52: 17-21.
- Kauffman H, Reddy APK, Hsieh SPY, Merca SD, Kauffman EJ. (1973) An improved technique for evaluating resistance of rice varieties to *Xanthomonas*

oryzae. Plant Dis Rep 57: 537-541.

- 35. Poland JA, Brown PJ, Sorrells ME, Jannink JL. (2012) Development of high-density genetic maps for barley and wheat using a novel two-enzyme genotyping-bysequencing approach (T. Yin, editor). PLoS One 7: e32253. doi: 10.1371/journal.pone.0032253.
- Furuta T, Ashikari M, JenaK K, Doi K, Reuscher S. (2017) Adapting genotyping-by-sequencing for rice F2 populations. Genes Genomes Genet 7: 881-893.
- 37. Glaubitz J, Casstevens T, Lu F, Harriman J, Elshire R, Sun Q, Buckler E. (2014) TASSEL-GBS: A high capacity genotyping by sequencing analysis pipeline. PLoS One 9. doi: 10.1371/journal.pone.0090346.
- 38. Swarts K, Li H, Alberto Romero Navarro J, An D, Romay M, Hearne S, Acharya C, Glaubitz J, Mitchell S, Elshire R, Buckler E, Bradbury P. (2014) Novel methods to optimize genotypic imputation for low-coverage, next-generation sequence data in crop plants. Plant Genome 7. doi: 10.3835/plantgenome2014.05.0023.
- 39. Ohsumi A, Takai T, Ida M, Yamamoto T, Arai-Sanoh Y, Yano M, Ando T, Kondo M. (2011) Evaluation of yield performance in rice near-isogenic lines with increased spikelet number. F Crop Res 120: 68-75.
- 40. Takai T, Matsuura S, Nishi T, Ohsumi A, Shiraiwa T, Horie T. (2006) Rice yield potential is closely related to crop growth rate during late reproductive period. Field Crop Res 96: 328-335.

収量関連遺伝子と白葉枯病抵抗性遺伝子の 戻し交配によるピラミディング系統の評価

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イネ収量関連遺伝子 WFPは、一次枝梗数増加による着粒数増加をもたらす。IR24に白葉枯病抵抗性遺伝子群(Xa4、xa5、 xa13、Xa21)を集積したIRBB60へ、ST-12が持つ WFPの導入を行った。IRBB60を反復親とした戻し交雑後代BC₂F₂と BC₂F₃世代で、DNAマーカーの利用と一次枝梗数を含む収量形質の評価から、3ピラミディング系統(PL-1、PL-4、PL-5) を選抜した。BC₂F₄において、これらはWFPによる一次枝梗数と着粒数の増加を示したが、4白葉枯病抵抗性遺伝子の集 積が不完全で、白葉枯病レース群への反応の差が見られた。しかし、PL-5はIRBB60よりも収量増加が見込まれ、今回供試 した白葉枯病レース(ベトナム、ミャンマー、インド由来6レース)の内5レースに対して抵抗性を示したことから、PL-5 は東南アジアの熱帯地域での展開を見込んだ候補系統となり得ると考えられた。

キーワード:イネ、WFP 遺伝子、白葉枯病抵抗性、ピラミディング、高収量性